

A PROPOSAL FOR CALCULATING THE NO-OBSERVED-ADVERSE-EFFECT LEVEL (NOAEL) FOR ORGANIC COMPOUNDS RESPONSIBLE FOR LIVER TOXICITY BASED ON THEIR PHYSICOCHEMICAL PROPERTIES

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Abstract

Objectives: Both environmental and occupational exposure limits are based on the no-observed-adverse-effect level (NOAEL), lowest-observed-adverse-effect level (LOAEL) or benchmark dose (BMD) deriving from epidemiological and experimental studies. The aim of this study is to investigate to what extent the NOAEL values for organic compounds responsible for liver toxicity calculated based on their physicochemical properties could be used for calculating occupational exposure limits. **Material and Methods:** The distribution coefficients from air to the liver ($\log K_{\text{liver}}$) were calculated according to the Abraham solvation equation. NOAEL and LOAEL values for early effects in the liver were obtained from the literature data. The descriptors for Abraham's equation were found for 59 compounds, which were divided into 2 groups: "non-reactive" (alcohols, ketones, esters, ethers, aromatic and aliphatic hydrocarbons, amides) and "possibly reactive" (aldehydes, allyl compounds, amines, benzyl halides, halogenated hydrocarbons, acrylates). **Results:** The correlation coefficients between $\log\text{-}\log K$ and \log NOAEL for non-reactive and reactive compounds amounted to $r = -0.8123$ and $r = -0.8045$, respectively, and were statistically significant. It appears that the Abraham equation could be used to predict the NOAEL values for compounds lacking information concerning their liver toxicity. **Conclusions:** In view of the tendency to limit animal testing procedures, the method proposed in this paper can improve the practice of setting exposure guidelines for the unstudied compounds.

Key words:

Liver, NOAEL, LOAEL, Organic compounds, Physicochemical properties, Abraham's Eq

INTRODUCTION

Both, environmental and occupational exposure limits for chemical substances are developed to prevent and control potential health hazards.

Generally, for all the effects with the exception of those induced by direct interaction of the compound or its

metabolites with genetic material, it is assumed that there is a threshold exposure, below which the probability of harm is negligible. Occupational Exposure Limits (OELs), Reference Doses (RfD) or Concentrations (RfC) are based on the points of departure such as the no-observed-adverse-effect level (NOAEL) or the lowest-observed-adverse-effect level

This project was conducted and financed under the EU 6th Framework Integrated Project OSIRIS (Contract No. GOCE-ET-2007-037017) (<http://osiris-reach.eu>). Project manager: Gerrit Schüürmann, Prof.

Received: October 11, 2013. Accepted: March 31, 2014.

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(LOAEL) deriving from industrial settings as well as experimental studies. In view of the scarcity of epidemiological data and the tendency to limit animal testing procedures or human volunteer studies, the possibility of calculating the above-mentioned points of departure for organic compounds on the basis of their physicochemical properties is of great importance.

There is a large number of steps on the pathway between the external dose administration and the final toxic effect. The continuous process between the external dose and the toxic response can be subdivided into steps related to the distribution of the chemical in the body (toxicokinetics) and those related to the actions of the chemical in the organism (toxicodynamics). Transfer of organic compounds from the air to the target tissues is of crucial significance in understanding their potential toxic effects. The processes involved in the absorption and distribution of xenobiotics are similar. They are presently described by physiologically based toxicokinetic (PBTK) models. However, such models are still scarce, complicated and developed only for single substances.

Abraham [1] constructed a general equation that seems satisfactory in terms of explaining the transfer of VOCs from the gaseous phase to a large number of condensed phases, including biophases (e.g. mucosa, serum for different organs).

$$SP = c + eE + sS + aA + bB + lL \quad (1)$$

In this equation, the dependent value SP is some free energy related property, such as an adsorption or absorption constant, $\log K$, where K is the gas to solvent partition coefficient for a series of VOCs into the given solvent or condensed phase, or a biological property of VOC, such as an odour or nasal pungency threshold (NPT), for a series of VOCs.

The independent variables in Equation 1 – E, S, A, B, and L – are the properties, or descriptors, of the VOC's, i.e., E – excess molar refraction, S – dipolarity/polarisability, A and B – overall or effective hydrogen bond

activity and basicity, respectively, L – ($\log L^{16}$) is defined through L^{16} , the VOC hexadecane-air partition coefficient at 298 K, and is a measure of VOC lipophilicity.

The coefficients c, e, s, a, b and l are found by multiple linear regression analysis. They reflect the complementary properties of the receptor phase. The e coefficient gives the tendency of the phase to interact with a compound through polarisability-type interactions, mostly via electron pairs. The s coefficient is a measure of the phase dipolarity/polarisability. The a coefficient represents the complementary property to the compound hydrogen-bond acidity, and so, is a measure of the phase hydrogen-bond basicity. Likewise, the b coefficient is a measure of the phase hydrogen-bond acidity. Finally, the l coefficient is a measure of the hydrophobicity of the phase.

In order to apply Equation 1 to any given process, a correlation between a reasonable number of data concerning the property is needed as the dependent variable. Simple multiple linear regression against the known compound descriptors leads to the coefficients. Once the latter have been calculated, then for any other compound for which the descriptors are available, the property can be calculated [2]. The descriptors for about 5000 compounds are available in the Pharma Algorithms database, ADME Boxes, Version 4.0, 2008. (Pharma Algorithms Inc., Toronto, ON, Canada). Additional descriptors, if required, can be predicted from the structure [3].

The Abraham's group have developed several equations enabling evaluation of the transfer of organic compounds to different organs and tissues such as liver [4], brain [5] or lungs [6].

Possible applications of the equations developed by Abraham et al. for prediction of retention of volatile organic compounds in the respiratory tract and for calculating occupational exposure limits for volatile organic compounds acting as sensory irritants have already been published [7,8].

The aim of this study is to investigate to what extent the relationship between the distribution coefficients K_{liver} from the air to the liver [4] could be used for calculating NOAEL values for organic compounds that are responsible for liver toxicity.

MATERIAL AND METHODS

Calculation of the distribution coefficients K_{liver}

Distribution coefficients K_{liver} from the air to the liver were calculated according to the Equation 2 published in the Pharma Algorithms database, ADME Boxes, Version 4.0, 2008. (Pharma Algorithms Inc., Toronto, ON, Canada):

$$\text{Log } K_{\text{liver}} = -1.031 + 0.059E + 0.774S + 0.593A + 1.049B + 0.654L \quad (2)$$

The descriptors E, S, A, B and L were also obtained from the Pharma Algorithms database.

No-observed-adverse-effect level (NOAEL) and LOAEL values were obtained from the literature data. Compounds which caused early non-neoplastic effects in the liver, as a result of inhalation or oral exposure, were considered. The increased liver weight was not considered as an adverse effect because it may reflect physiological adaptation to the presence of particular substance. The Abraham's descriptors have been found for 59 compounds.

Abraham's equation describes the transfer of compounds to the liver regardless of their toxicodynamics. Biological activity of a chemical depends on the transport from the site of administration to the site of action and reaction of the unchanged compound or its metabolites with the receptor or target (i.e., biological activity is a function of partition and reactivity). If a structure/activity model is deficient in modelling either partition or reactivity, only a partial correlation with the biological response is likely to be observed [9].

Since it was impossible to evaluate the balance of local bioactivation and detoxication of particular compounds as well as the possible interaction of unchanged compounds and their metabolites with intracellular targets, all compounds were divided into 2 groups: "non-reactive" (alcohols, ketones, esters, ethers, aromatic and aliphatic hydrocarbons, amides) and "possibly reactive" (aldehydes, allyl compounds, amines, benzyl halides, halogenated hydrocarbons, acrylates) [10].

The NOAEL and LOAEL values are presented in Tables 1 and 2.

To obtain the corresponding NOAEL values the LOAEL values were divided by 2 [64]. Then NOAELs obtained both as a result of inhalation and oral exposure in animals have been transformed into concentrations in mg/m^3 for workers according to the procedure described in the REACH document [65].

In the first step oral NOAEL for a given animal was transferred to humans with a factor of 4 for allometric scaling for rats, 7 for mice and 2.5 for other interspecies differences.

Using a standard human body weight (70 kg) and a default human breathing volume of 10 m^3 for workers in 8 h and light activity, this dose was then translated into the air concentration (NAEC) according to the equation:

$$\text{NAEC}_{\text{worker 8h}} = \text{NOAEL} \times 70 \text{ kg/allometric scaling factor} \times 10 \text{ m}^3 \quad (3)$$

Where inhalative data are concerned, air concentrations for animal and human exposure are generally compared directly. For workers the air concentration was corrected for the lung ventilation (6.7 m^3 for base level and 10 m^3 for light activity during 8 h):

$$\text{NAEC}_{\text{worker 8h}} = \text{NOAEL}_{\text{inh}} \times 6.7 \text{ m}^3/10 \text{ m}^3 \quad (4)$$

Statistical analysis

Statistical analysis was carried out using SPSS version 14. Linear regression analysis was applied to examine the correlation.

Table 1. NOAEL/LOAEL values for non-reactive compounds

Compound	CAS No.	Critical study [references]	NOAEL/ LOAEL*	NOAEL modified (mg/m ³)	Log NOAEL** modified	log-logK
Acetone	67-64-1	rats – gavage (30–90 days), an increased liver weight and liver-to-body weight ratio [11]	NOAEL: 2 500 mg/kg/day	4 375	3.64	0.055
Tert-Amyl methyl ether	994-05-8	rats, dogs – inhalation (6 h/day, 5 days/week, 90 day), an increased absolute and relative liver weights [12]	NOAEL: 4 180 mg/m ³	2 800	3.44	0.120
2-Butoxyethanol	11-76-2	rats, mice – inhalation (6 h/day, 5 days/week, 14 weeks), an increased number of liver Kupffer cells [13]	LOAEL: 302 mg/m ³ NOAEL: 151 mg/m ³	101	2.00	0.480
Cumene	98-82-8	rats – inhalation (6 h/day, 5 days/week, 13 weeks), an increased liver weight [14]	NOAEL: 2 460 mg/m ³	1 648	3.22	0.390
Cyclohexane	110-82-7	rats – inhalation (6 h/day, 5 days/week, 90 days), an increased liver weight, centrilobular hypertrophy [15]	LOAEL: 24 500 mg/m ³ NOAEL: 12 250 mg/m ³	7 350	3.86	0.120
Cyclohexanol	108-93-0	rabbits – inhalation (6 h/day, 5 days/week, 5–11 weeks), slight degenerative changes in the liver [16]	LOAEL: 593 mg/m ³ NOAEL: 269 mg/m ³	180	2.26	0.370
Cyclohexanone	108-94-1	rabbits – inhalation (6 h/day, 50 days), barely demonstrable degenerative changes in the liver [16]	LOAEL: 762 mg/m ³ NOAEL: 381 mg/m ³	255	2.41	0.370
<i>N,N</i> -Dimethylacetamide	127-19-5	rats – inhalation (6 h/day, 5 days/week, 6 months), focal necrosis of liver [17]	LOAEL: 694 mg/m ³ NOAEL: 347 mg/m ³	233	2.37	0.410
Dimethylformamide	68-12-2	rats – inhalation (6 h/day, 10 days), slightly enlarged livers [18]	NOAEL: 272 mg/m ³	182	2.26	0.370
Ethanol	64-17-5	rats – gavage (12 weeks), a fatty liver and necrosis [19]	LOAEL: 10 000 mg/kg/day NOAEL: 5 000 mg/kg/day	4 375 8 750	3.94	0.060
Ethyl acetate	141-78-6	rabbits – inhalation (65 four-hour exposures), liver damage [20]	LOAEL: 16 020 mg/m ³ NOAEL: 8 010 mg/m ³	3 601	3.55	0.130
Ethyl benzene	100-41-4	rats – inhalation (7 h/day, 5 day/week, 6 months), a cloudy swelling in the liver [21]	LOAEL: 1 736 mg/m ³ NOAEL: 868 mg/m ³	582	2.76	0.340

Table 1. NOAEL/LOAEL values for non-reactive compounds – cont.

Compound	CAS No.	Critical study [references]	NOAEL/ LOAEL*	NOAEL modified (mg/m ³)	Log NOAEL** modified	log-logK
Ethyl tert-Butyl ether	637-92-3	rats – inhalation (6 h/day, 5 days/week, 4 weeks), a relative liver weight increase [12]	NOAEL: 8 360 mg/m ³	5 601	3.74	0.21
Indene	95-13-6	rats – inhalation (six 7.5-hour exposure periods), from slightly fatty degeneration to severe necrosis in the liver [22]	LOAEL: 3 792 mg/m ³ NOAEL: 1 896 mg/m ³	1 272	3.10	0.45
Methyl tert-butyl ether	1634-04-4	rats – inhalation (6 h/day, 5 days/week, 13 weeks), an increased liver weight [23]	NOAEL: 14 400 mg/m ³	9 648	3.98	0.02
Methylcyclohexane (cyclohexylmethane)	108-87-2	rabbits – inhalation (6 h/day, 90 h), a slight cellular liver injury [16]	LOAEL: 11 520 mg/m ³ NOAEL: 5 760 mg/m ³	3 865	3.59	0.16
Methyl isoamyl ketone	110-12-3	rats – inhalation (6 h/day, 5 days/week, 96 days), slight increases in liver weight [24]	NOAEL: 4 660 mg/m ³	3 122	3.49	0.38
Methyl isobutyl ketone	108-10-1	rats – inhalation (24 h/day, 2 weeks), an increased absolute liver weight and an increased organ-to-body-weight ratio [25]	NOAEL: 818 mg/m ³	548	2.73	0.30
Xylene (all isomers)	1330-20-7	rats – inhalation (8 h/day, 6 days/week, 110–130 days), a slight liver congestion [26]	LOAEL: 2 967 mg/m ³ NOAEL: 1 484 mg/m ³	994	2.99	0.33

* NOAEL – no-observed-adverse-effect level.

** LOAEL – lowest-observed and adverse-effect level.

Table 2. NOAEL/LOAEL values for reactive compounds

Compound	CAS No	Critical study [references]	NOAEL/ LOAEL*	NOAEL modified (mg/m ³)	Log NOAEL** modified	log-logK
Acetonitrile	1975-05-08	rats – inhalation (6 h/day, 5 days/week, 13 weeks), an increased liver weight [27]	NOAEL: 2 688 mg/m ³	1 800.0	3.25	-0.140
Aldrin	309-00-2	rats – diet (2 days), an increased liver weight [28]	NOAEL: 1.2 mg/kg/day	2.1	0.32	0.822
Allyl alcohol	107-18-6	rats – drinking water (15 weeks), hepatotoxicity [29]	LOAEL: 17 mg/kg/day NOAEL: 8.5 mg/kg/day	14.8	1.17	0.160
Allyl chloride	107-05-1	rats, mice – inhalation (6 h/day, 5 days/week, 3 months), no changes in liver attributable to allyl chloride [30]	NOAEL: 787 mg/m ³	527.0	2.72	-0.250

Table 2. NOAEL/LOAEL values for reactive compounds – cont.

Compound	CAS No	Critical study [references]	NOAEL/ LOAEL*	NOAEL modified (mg/m ³)	Log NOAEL** modified	log-logK
Bromoform	75-25-2	rats – diet (30 days), liver swelling [31]	LOAEL: 56 mg/kg NOAEL: 28 mg/kg	49.00	1.69	0.33
Carbon tetrachloride	56-23-5	rats, mice – inhalation (6 h/day, 5 days/week, 13 weeks), an increased liver weight, fatty changes [32]	LOAEL: 63 mg/m ³ NOAEL: 31.5 mg/m ³	21.10	1.33	0.05
Chlorobenzene	108-90-7	rats – inhalation (7 h/day, 5 days/week, 24 weeks), an increased liver weight, congestion [33]	LOAEL: 346 mg/m ³ NOAEL: 173 mg/m ³	115.00	2.06	0.30
Chloroform	67-66-3	rats – inhalation (7 h/day, 5 days/week, 6 months), a centrilobular granular degeneration [34]	LOAEL: 243 mg/m ³ NOAEL: 121 mg/m ³	81.50	1.91	0.09
Cyclonite	121-82-4	rats – diet (90 days), an enlarged liver [35]	NOAEL: 30 mg/kg/day	52.50	1.72	0.87
o-Dichlorobenzene	95-50-1	rats – gavage (138 doses), an increased liver weight [36]	NOAEL: 188 mg/kg	329.00	2.52	0.42
p-Dichlorobenzene	106-46-7	rats – diet (130 oral doses 192 days), an increased liver weight [37]	NOAEL: 188 mg/kg	3.10	1.90	0.38
1,1-Dichloroethane	75-34-3	rats, guinea pigs, rabbits – inhalation (6 h/day, 5 days/week, 13 weeks) [38]	NOAEL: 2 025 mg/m ³	1 956.00	3.29	-0.21
Dichloromethane	75-09-2	dogs, guinea pigs – inhalation (4 h/day, 5 days/week, 7.5 weeks), a moderate fatty liver degeneration [39]	LOAEL: 35 300 mg/m ³ NOAEL: 17 650 mg/m ³	11 825.00	4.07	-0.45
Diethanolamine	111-42-2	rats – inhalation (6 h/day, 5 days/week, 13 weeks), an increased liver weight [40]	NOAEL: 29 mg/m ³	19.40	1.29	0.60
Diethylamine	109-89-7	rabbits – inhalation (7 h/day, 5 days/week, 6 weeks), a parenchymatous liver degeneration with evidence of cell regeneration [41]	LOAEL: 105 mg/m ³ NOAEL: 52.5 mg/m ³	35.20	1.55	0.25
Diphenylamine	122-39-4	beagle dogs – diet (2 years), a peripherolobular fatty change with a marked increase in liver weight [42]	LOAEL: 110 mg/kg/day NOAEL: 55 mg/kg/day	192.00	2.28	0.68
Endosulfan	115-29-7	rats – diet (5 mg/kg, 30 days), an increased liver weight [43]	NOAEL: 5 mg/kg	8.75	0.94	1.05

Table 2. NOAEL/LOAEL values for reactive compounds – cont.

Compound	CAS No	Critical study [references]	NOAEL/ LOAEL*	NOAEL modified (mg/m ³)	Log NOAEL** modified	log-logK
Ethyl bromide	74-96-4	rats – inhalation (4 h/day, 6 months), hepatic function disruption [44]	LOAEL: 2 410 mg/m ³ NOAEL: 1 205 mg/m ³	807.00	2.91	-0.21
Ethyl chloride	75-00-3	rats – inhalation (6 h/day, 5 days/week, 13 weeks), an increased liver to body weight ratios [45]	NOAEL: 50 540 mg/m ³	33 862.00	4.53	-0.72
Ethylene dichloride	107-06-2	guinea pigs – inhalation (7 h/day, 5 days/week, 226 days), an increased liver weight [46]	NOAEL: 405 mg/m ³	271.00	2.43	-0.01
Halothane	151-67-7	rats, rabbits – inhalation (5 h/day, 5 days/week, 7 weeks), a centrilobular fatty liver infiltration [47]	LOAEL: 4 030 mg/m ³ NOAEL: 2 015 mg/m ³	1 350.00	3.13	-0.10
Heptachlor	76-44-8	rats – in diet (80 days), hepatic necrosis and synthesis of smooth endoplasmic reticulum [47]	LOAEL: 2 mg/kg/day NOAEL: 1 mg/kg/day	1.75	0.24	0.87
Hexachlorobenzene	118-74-1	rats – in diet (21 days), a relative liver weight increase, a centrilobular increase in hepatocyte size [48]	LOAEL: 8 mg/kg/day NOAEL: 4 mg/kg/day	7.00	0.84	0.63
Hexachlorobutadiene	87-68-3	rats – in diet (30 days), minimal hepatocellular swelling [49]	LOAEL: 100 mg/kg NOAEL: 50 mg/kg	87.50	1.94	0.53
Hexachloroethane	67-72-1	rats – diet (16 weeks), an increased liver weight, slight swelling of hepatocytes [50]	LOAEL: 62 mg/kg/day NOAEL: 31 mg/kg/day	54.20	1.73	0.44
Methyl chloroform	71-55-6	rats, guinea pigs – inhalation (1.2–3 h/day, 3 months), pathologic changes in liver [51]	LOAEL: 5 440 mg/m ³ NOAEL: 2 720 mg/m ³	1 842.00	3.26	-0.05
4,4'-Methylene dianiline	101-77-9	rats – oral intubation (16 weeks), a minor liver damage [52]	LOAEL: 8 mg/kg/day NOAEL: 4 mg/kg/day	7.00	0.85	0.88
Picloram	1918-02-1	rats – food (2 years), an increased liver weight, a centrilobular hepatocellular hypertrophy [53]	LOAEL: 60 mg/kg/day NOAEL: 30 mg/kg/day	53.00	1.72	0.85

Table 2. NOAEL/LOAEL values for reactive compounds – cont.

Compound	CAS No	Critical study [references]	NOAEL/ LOAEL*	NOAEL modified (mg/m ³)	Log NOAEL** modified	log-logK
Propargyl alcohol	107-19-7	rats – inhalation (7 h/day, 5 days/week, 3 months), an increased liver weight [19]	NOAEL: 184 mg/m ³	123.00	2.09	0.140
Propylene dichloride	78-87-5	rats – in diet (5 times weekly, 13 weeks), degenerative effects on the centrilobular hepatocytes [54]	LOAEL: 250 mg/kg/day NOAEL: 125 mg/kg/day	218.00	2.34	0.069
Pyridine	110-86-1	rats – diet (90 days), mild hepatotoxicity, inflammatory lesions, increased liver and liver to body weight ratios [55]	LOAEL: 25 mg/kg/day NOAEL: 12.5 mg/kg/day	21.90	1.34	0.310
1,1,2,2-Tetrachloroethane	79-34-5	rats, rabbits – inhalation (3–4 h/day, 11 months), a liver damage [50]	LOAEL: 100 mg/m ³ NOAEL: 50 mg/m ³	33.50	1.53	0.360
Tetrachloroethylene	127-18-4	mice – lavage (5 days/week, 6 weeks), an increased liver weight and triglycerides [56]	LOAEL: 100 mg/kg/day NOAEL: 50 mg/kg/day	50.00	1.70	0.310
1,2,4-Trichlorobenzene	120-82-1	rats – inhalation (7 h/day, 5 days/week, 26 weeks), microscopic changes in liver parenchyma after 4 or 13 weeks of exposure [57]	LOAEL: 185 mg/m ³ NOAEL: 92.5 mg/m ³	62.00	1.79	0.460
1,1,2-Trichloroethane	79-00-5	mice – drinking water (90 day), clinical chemistry indications of adverse effects on the liver [58]	LOAEL: 305 mg/kg/day NOAEL: 153 mg/kg/day	153.00	2.18	0.170
Trichloroethylene	79-01-6	rats – inhalation (7 h/day, 5 days/week, 6 months), an increased liver weight [59]	NOAEL: 2 154 mg/m ³	1 443.00	3.16	0.110
1,2,3-Trichloropropane	96-18-4	rats – inhalation (6 h/day, 5 days/week, 13 weeks), an increased liver weight [60]	NOAEL: 30 mg/m ³	20.10	1.30	0.340
2,4,6-Trinitrotoluene (TNT)	118-96-7	dogs – gelatin capsules (0.5 mg/kg/day, 6 months), frank hepatotoxicity [61]	LOAEL: 0.5 mg/kg/day NOAEL: 0.25 mg/kg/day	0.87	-0.06	0.800
Vinylidene chloride	75-35-4	rats, rabbits, guinea pigs – inhalation (8 h/day, 5 days/week, 6 months), some hepatic degeneration [62]	LOAEL: 100 mg/m ³ NOAEL: 50 mg/m ³	34.00	1.53	-0.110
Xylidine	1300-73-8	rats – gavage (4 weeks), an increased liver weight [63]	NOAEL: 20 mg/kg	35.00	1.54	0.570

Abbreviations as in Table 1.

RESULTS AND DISCUSSION

The NOAEL values for non-reactive and reactive compounds as well as the respective $\log\text{-}\log K_{\text{liver}}$ values are presented in Tables 1 and 2.

The obtained relationships between $\log\text{-}\log K$ and NOAEL values for non-reactive and reactive compounds are presented on Figures 1 and 2. The correlation coefficients between $\log\text{-}\log K$ and \log NOAEL for non-reactive (Figure 1) and reactive compounds (Figure 2) amounted to $r = -0.8045$ and $r = -0.8123$, respectively, and were statistically significant.

The method for calculation of the NOAEL values was validated against the already established OEL values. For this purpose, LOAELs for 17 compounds, where the liver was considered as the critical organ for setting OELs by ACGIH [50], were calculated on the basis of their $\log\text{-}\log K_{\text{liver}}$ values according to the regression equations for reactive and non-reactive compounds. To obtain the OELs, the calculated NOAELs were divided by 12.5 according to the recommendations provided in the REACH document [65] (assessment factors: 2.5 for interspecies differences other than factor for allometric scaling and 5 for intraspecies differences) (Table 3).

Correlation between the mean OEL values published by at least 2 out of 3 organizations [50,66,67] and those calculated ones was very high ($r = 0.897$, $p < 0.000$) (Figure 3). Ratio of the calculated values to the mean OELs depends on the assumed assessment factors. Using the factor

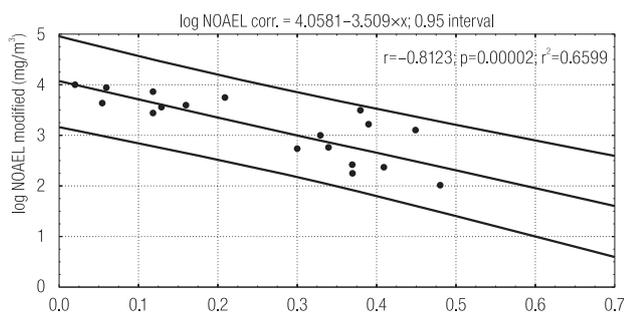


Fig. 1. Relationship between the $\log\text{-}\log K$ and \log NOAEL – non-reactive compounds

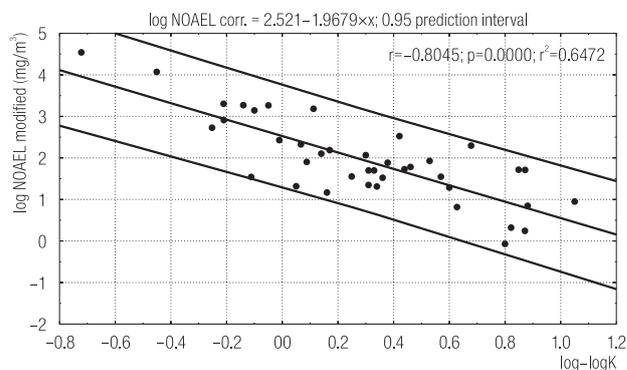


Fig. 2. Relationship between the $\log\text{-}\log K$ and \log NOAEL – reactive compounds

of 12.5, according to the REACH document [65], 7 of the calculated OELs were lower and 10 were higher than the mean published values. However, taking into account the divergences between the OELs in different countries, the possibility of practical usage of the proposed way of calculating LOAEL values seems very promising. According to the Setubal-principles developed by the Expert Group of the OECD Work Program on QSARs, the model for regulatory purposes should be associated with the following information: a defined endpoint, an unambiguous algorithm, a defined domain of applicability, appropriate measures of goodness-of-fit, robustness and predictivity

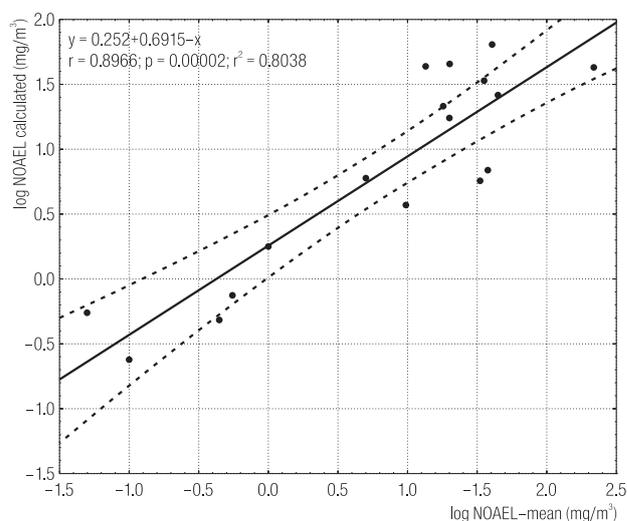


Fig. 3. Relationship between the mean and the calculated NOAEL values

Table 3. Comparison between the published Occupational Exposure Limits (OELs) and the calculated values

Substances	Published OELs (mg/m ³)					Calculated OELs	Log
	TLV	MAK	NDS	M	Log-x		
Bromoform*	5.20	–	5.00	5.10	0.70	5.93	0.77
Carbon tetrachloride*	31.00	3.20	20.00	18.10	1.26	21.00	1.32
Chlorobenzene*	46.00	47.00	23.00	38.70	1.58	6.80	0.83
Chloroform*	49.00	2.50	8.00	19.80	1.30	17.50	1.24
Diethanoloamine*	1.00	1.00	–	1.00	0.00	1.75	0.24
N,N-Dimethyl acetamide**	36.00	36.00	35.00	35.60	1.55	33.30	1.52
Dimethylformamide**	30.00	15.00	15.00	20.00	1.30	45.00	1.65
Endosulfan*	0.10	–	0.10	0.10	–1.00	0.23	–0.63
Ethyl bromide*	22.00	50.00	50.00	40.60	1.61	68.70	1.80
Ethylene dichloride*	40.00	–	50.00	45.00	1.65	25.90	1.41
Halothane*	404.00	41.00	–	222.00	2.34	41.80	1.62
Heptachlor*	0.05	0.05	–	0.05	–1.30	0.53	–0.27
Hexachloro ethanol*	9.70	9.80	10.00	9.80	0.99	3.60	0.56
4,4'-Methylene dianiline*	0.81	–	0.08	0.45	–0.35	0.48	–0.32
Trichloropropane*	60.00	–	7.00	33.50	1.52	5.66	0.75
Trinitrotoluene*	0.10	–	1.00	0.55	–0.26	0.72	–0.14
Vinylidene chloride*	20.00	8.00	12.50	13.50	1.13	42.80	1.63

TLV – ACGIH – USA – threshold limit value – American Conference of Governmental Industrial Hygienists [50].

MAK – DFG – Germany – Maximum Concentrations – Deutsche Forschungsgemeinschaft [66].

NDS (MAC) – CIOP-PIB – maximum admissible concentrations time weighed [67].

OEL – occupational exposure limits.

M – model at the prediction of safe OELs.

* Reactive compounds.

** Non-reactive compounds.

and a mechanistic interpretation, if possible [68]. In the proposed model, aiming at the prediction of safe OELs, the first 4 points have been fulfilled.

About 45% of the Threshold Limit Values (TLV), established by the American Conference of Governmental Industrial Hygienists, for organic compounds are based on the sensory irritation effect. Liver is considered to be a critical organ for 13%, CNS for 12% of substances. Other effects include inhibition of the acetylcholinesterase activity, methaemoglobinemia, and other, mainly in a form of cancer and reproductive toxicity. In total,

about 70% of TLV's are based on sensory irritation and disturbances in the liver or CNS functions [50].

The results showing the possibility of OELs prediction based on sensory irritation by means of the same general algorithm have already been published [8]. It is still necessary to verify the possibility of application of the same general algorithm in the case of the central nervous system [5]. If the results are positive, then it would be possible to predict safe occupational exposure limits for the majority of compounds, which can be potentially used in industry, except for the compounds

responsible for effects such as cancer and reproductive toxicity, inhibition of acetylcholinesterase activity or methaemoglobinemia.

CONCLUSIONS

The obtained results suggest that NOAELs and consequently OELs for organic compounds responsible for liver toxicity can be predicted based on the $\log K_{\text{liver}}$ values calculated according to the Abraham's Eq. Also validation of the proposed method gave satisfactory results. Correlation between the mean OEL values published by at least 2 out of 3 organizations [50,66,67] and those calculated ones was very high ($r = 0.897$, $p < 0.000$). In view of the scarcity of human data and the tendency to limit animal testing procedures, the method proposed in this paper can improve the practice of setting exposure guidelines for the unstudied compounds.

In the presented publication the Abraham equation gives positive results. Please remember that the equation refers only to the toxicokinetics. The results are valid only for the toxicokinetic characteristics of the chemical compound, i.e. they depend solely on the speed of their deposition in a specified system or organ. In the future, some basic correction to account for the reactivity of individual toxicodynamics of the compounds may be necessary.

REFERENCES

1. Abraham MH. Scales of solute hydrogen-bonding: Their construction and application of physicochemical and biochemical processes. *Chem Soc Rev.* 1993;22:73–83, <http://dx.doi.org/10.1039/cs9932200073>.
2. Abraham MH, Gola JMR, Cometto-Muniz JE, Cain WS. The correlation and prediction of VOC threshold for nasal pungency, eye irritation and odour in humans. *Indoor Built Environ.* 2001;10:252–7, <http://dx.doi.org/10.1177/1420326X0101000320>.
3. Abraham MH, Ibrahim A, Acree Jr WE. Air to blood distribution of volatile organic compounds: A linear free energy analysis. *Chem Res Toxicol.* 2005;18:904–11, <http://dx.doi.org/10.1021/tx050066d>.
4. Abraham MH, Ibrahim A, Acree Jr WE. Air to liver partition coefficients for volatile organic compounds and blood to liver partition coefficients for volatile organic compounds and drugs. *Eur J Med Chem.* 2007;42:743–51, <http://dx.doi.org/10.1016/j.ejmech.2006.12.011>.
5. Abraham MH, Ibrahim A, Acree Jr WE. Air to brain, blood to brain and plasma to brain distribution of volatile organic compounds: Linear free energy analyses. *Eur J Med Chem.* 2006;41:494–502.
6. Abraham MH, Ibrahim A, Acree Jr WE. Air to lung partition coefficients for volatile organic compounds and blood to lung partition coefficients for volatile organic compounds and drugs. *Eur J Med Chem.* 2008;43:478–85, <http://dx.doi.org/10.1016/j.ejmech.2007.04.002>.
7. Jakubowski M, Czerczak S. Calculation of retention of volatile organic compounds in the lung on the basis of their physicochemical properties. *Environ Toxicol Pharmacol.* 2009;2:311–5, <http://dx.doi.org/10.1016/j.etap.2009.05.011>.
8. Jakubowski M, Czerczak S. A proposal for calculating occupational exposure limits for volatile organic compounds acting as sensory irritants on the basis of their physicochemical properties. *J Occup Environ Hyg.* 2010;7:429–34, <http://dx.doi.org/10.1080/15459624.2010.483983>.
9. Barrat MD, Rodford RA. The computational prediction of toxicity. *Curr Opin Chem Biol.* 2001;5:383–8, [http://dx.doi.org/10.1016/S1367-5931\(00\)00218-0](http://dx.doi.org/10.1016/S1367-5931(00)00218-0).
10. Abraham MH, Kumarsingh R, Cometto-Muniz JE, Cain WS. An algorithm for nasal pungency thresholds in man. *Arch Toxicol.* 1998;72:227–32, <http://dx.doi.org/10.1007/s002040050493>.
11. EPA U.S. Environmental Protection Agency. Ninety-day gavage study in albino rats using acetone. Washington, DC: U.S. EPA, Office of Solid Waste; 1986.

12. White RD, Daughtrey WC, Wells MS. Health effects of inhaled tertiary amyl methyl ether and tertiary butyl ether. *Toxicol Lett.* 1995;82/83:719–24, [http://dx.doi.org/10.1016/0378-4274\(95\)03590-7](http://dx.doi.org/10.1016/0378-4274(95)03590-7).
13. NTP, U.S. National Toxicology Program. Toxicology and carcinogenesis studies of 2-butoxyethanol (CAS NO 111-76-2) in F344/N rats and B6C3F1 mice (inhalation studies). TR-484. Research Triangle Park: NTP; 2000 [cited 2000 March 17]. Available from: http://ntp.niehs.nih.gov/ntp/htdocs/lt_rpts/tr484.pdf.
14. Cushman JR, Norris JC, Dodd DE, Darmer KI, Morris CR. Subchronic inhalation toxicity and neurotoxicity assessment of cumene in fischer 344 rats. *J Am Coll Toxicol.* 1995;14:129–47, <http://dx.doi.org/10.3109/10915819509008687>.
15. Malley LA, Bamberger JR, Stadler JC, Elliot GS, Hansen JF, Chiu T, et al. Subchronic toxicity of cyclohexane in rats and mice by inhalation exposure. *Drug Chem Tox.* 2000;23(4):513–37, <http://dx.doi.org/10.1081/DCT-100101969>.
16. Treon JF, Crutchfield WE, Kitzmiller KV. The physiological response of animals to cyclohexane, methylcyclohexane, and certain derivatives of these compounds. II. Inhalation. *J Ind Hyg Toxicol.* 1943;25:323–47.
17. Horn HJ. Toxicology of dimethylacetamide. *Toxicol Appl Pharmacol.* 1961;3:12–24, [http://dx.doi.org/10.1016/0041-008X\(61\)90003-5](http://dx.doi.org/10.1016/0041-008X(61)90003-5).
18. Clayton JW Jr, Barnes JR, Hood DB, Schepers GWH. The inhalation toxicity of dimethylformamide (DMF). *Am Ind Hyg Assoc J.* 1963;24:144–54, <http://dx.doi.org/10.1080/00028896309342942>.
19. Bevan C. Monohydric alcohols-C1 to C6. In: Bingham E, Cohns B, Powell CH, editors. *Patty's Toxicology*. 5th ed. Vol. 6. New York: John Wiley&Sons; 2001. p. 365–541, <http://dx.doi.org/10.1002/0471435139.tox077>.
20. Hathaway GJ, Proctor NH, Hughes JP, editors. Ethyl acetate. In: Hathaway GJ, Proctor NH, Hughes JP, editors. *Proctor and Hughes' chemical hazards of the workplace*. 4th ed. New York: Van Nostrand Reinhold; 1996. p. 306.
21. Wolf MA, Rowe VK, McCollister DD. Toxicological studies of certain alkylated benzenes and benzene. *Arch Ind Health.* 1956;14:387–98.
22. Cameron GR, Doniger CR. The toxicity of indene. *J Pathol Bacteriol.* 1939;49:529–33, <http://dx.doi.org/10.1002/path.1700490308>.
23. Lington AW, Dodd DF, Ridlon SA, Douglas JF, Kneiss JJ, Andrews LS. Evaluation of 13-week inhalation toxicity study on MTBE in Fisher 344 rats. *J Appl Toxicol.* 1997;17 (Suppl 1):S37–44, [http://dx.doi.org/10.1002/\(SICI\)1099-1263\(199705\)17:1+%3CS37::AID-JAT409%3E3.3.CO;2-H](http://dx.doi.org/10.1002/(SICI)1099-1263(199705)17:1+%3CS37::AID-JAT409%3E3.3.CO;2-H).
24. Katz GV, Renner ER, Terhaar CJ. Subchronic inhalation toxicity of methyl isoamyl ketone in rats. *Fund Appl Toxicol.* 1986;6:498–505, [http://dx.doi.org/10.1016/0272-0590\(86\)90223-X](http://dx.doi.org/10.1016/0272-0590(86)90223-X).
25. MacEwen JD, Vernot EH, Haun CC. Effect of 90-day continuous exposure to methylisobutylketone on dogs, monkeys, and rabbits. Springfield: National Technical Information Service; 1971.
26. Fabre R, Truhaut R, Laham S. Toxicological research on replacement solvents for benzene. IV. Study on xylenes. *Arch Mal Prof.* 1960;21:301–13.
27. NTP U.S. National Toxicology Program. Toxicology and carcinogenesis studies of acrylonitrile. Report TR 447. Research Triangle Park: NTP; 1996 [cited 2013 Sept 12]. Available from: http://ntp.niehs.nih.gov/ntp/htdocs/lt_rpts/tr447.pdf.
28. Cleveland FP. A summary of work on aldrin and dieldrin toxicity at the Kettering Laboratory. *Arch Environ Health.* 1966;13:195–8, <http://dx.doi.org/10.1080/00039896.1966.10664532>.
29. Carpanigni FMB, Gaunt IF, Hardy J, Gangolli SD, Butterworth KR, Lloyd AG. Short-term toxicity of allyl alcohol in rats. *Toxicology.* 1978;9:29–45, [http://dx.doi.org/10.1016/0300-483X\(78\)90029-X](http://dx.doi.org/10.1016/0300-483X(78)90029-X).
30. Quast JF, Henck JW, Schuetz DJ. Allyl chloride-subchronic studies Results of an inhalation 4-day probe and 90-day subchronic study in laboratory rodents. Final Report. U.S.A, Midland: Toxicology Research Laboratory, Dow Chemical; 1982.

31. Aida Y, Takada K, Uchida O, Yasuhara K, Kurosawa Y, Tobe M. Toxicities of microencapsuled tribromomethane, dibromochloromethane and bromodichloromethane administered in the diet to Wistar rats for one month. *J Toxicol Sci.* 1992;17:119–33, <http://dx.doi.org/10.2131/jts.17.119>.
32. Nagano K, Umeda Y, Saito MI, Nishizawa T, Ikawa N, Aratio H, et al. Thirteen-week inhalation toxicity of carbon tetrachloride in rats and mice. *J Occup Health.* 2007;49:249–59, <http://dx.doi.org/10.1539/joh.49.249>.
33. Dilley JV, Lewis TR. Toxic evaluation of inhaled chlorobenzene. *Toxicol Appl Pharmacol.* 1978;45:327.
34. Torkelson TR, Oyen F, Rowe VK. The toxicity of chloroform as determined by single and repeated exposure of laboratory animals. *Am Ind Hyg Assoc J.* 1976;37:697–704, <http://dx.doi.org/10.1080/0002889768507551>.
35. Levine BS, Furedu EM, Gordon DE, Burns JM, Lish PM. Thirteen-week toxicity study of hexahydro-1,3,5-trinitro-1,3,5-triazine (RDX) in Fisher 344 rats. *Toxicol Lett.* 1981;8:241–5, [http://dx.doi.org/10.1016/0378-4274\(81\)90108-9](http://dx.doi.org/10.1016/0378-4274(81)90108-9).
36. Hollingsworth RL, Rowe VK, Oyen T, Torkelson MS, Adams EM. Toxicity of o-dichlorobenzene studies on animals and industrial experience. *AMA Arch Ind Health.* 1958;17:180–7.
37. Hollingsworth RL, Rowe VK, Oyen T, Hoyle HR, Spencer HC. Toxicity of paradichlorobenzene. *Arch Ind Health.* 1956;14:138–47.
38. Hofmann HY, Birnsteil H, Jobst P. On the inhalation toxicity of 1,1- and 1,2-Dichloroethane. *Arch Toxicol.* 1971;27:248–65, <http://dx.doi.org/10.1007/BF00315048>.
39. Heppel LA, Neal PA, Perrin TL. Toxicology of dichloromethane (Methylene chloride). I. Studies on toxicology of daily inhalation. *J Ind Hyg Toxicol.* 1944;26:8–16.
40. Beyer KHJr, Bergfeld WF, Berndt WO. Final report on the safety assessment of triethanolamine, diethanolamine, and monoethanolamine. *J Am Coll Toxicol.* 1983;2:183–235, <http://dx.doi.org/10.3109/10915818309142006>.
41. Brieger H, Hodes WA. Toxic effects of exposure to vapors of aliphatic amines. *AMA Arch Ind Hyg Occup Med.* 1986;3:287–91.
42. Thomas JO, Ribelin WE, Woodward JR, DeEds F. The chronic toxicity of diphenylamine for dogs. *Toxicol Appl Pharmacol.* 1967;11:184–94, [http://dx.doi.org/10.1016/0041-008X\(67\)90037-3](http://dx.doi.org/10.1016/0041-008X(67)90037-3).
43. Dikshith TSS, Raizada RB, Srivastava MK, Kaphalia BS. Response of rats to repeated oral administration of endosulfan. *Ind Health.* 1984;22:295–304, <http://dx.doi.org/10.2486/indhealth.22.295>.
44. Karimullina NK, Gizatullina AA. Effect of ethyl bromide on the liver. *Pharmacol Toxicol.* 1969;32:165–7.
45. NTP U.S. National Toxicology Program. Toxicology and carcinogenesis studies of chloroethane (ethyl chloride) in F344/N rats and B6C3F1 mice (inhalation studies). Technical Report 346. Research Triangle Park: NTP; 1989 [cited 2013 Sept 12]. Available from: http://ntp.niehs.nih.gov/ntp/htdocs/lt_rpts/tr346.pdf.
46. Spencer HC, Rowe VK, Adams EM. Vapor toxicity of ethylene dichloride determined by experiments on laboratory animals. *Arch Ind Hyg Occup Med.* 1951;4:482–93.
47. Chenoweth MB, Leong BKJ, Sparschu GL, Torkelson TR. Toxicity of methoxyflurane, halothane, and diethyl ether in laboratory animals on repeated inhalation of subanesthetic concentrations. In: Donald W, Benson MD. *Cellular Biology and Toxicity of Anesthetics*. Baltimore: Williams & Wilkins; 1972. p. 275–85.
48. International Agency for Research on Cancer (IARC). IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans, Vol. 20, Heptachlor and Heptachlor Epoxide. Lyon: IARC; 1979. p. 129–54.
49. Kuiper-Goodman T, Grant DL, Moodie CA, Korsrud GO, Munro IC. Subacute toxicity of hexachlorobenzene in the rat. *Toxicol Appl Pharmacol.* 1977;40:529–49, [http://dx.doi.org/10.1016/0041-008X\(77\)90078-3](http://dx.doi.org/10.1016/0041-008X(77)90078-3).
50. American Conference of Governmental Industrial Hygienists (ACGIH). 2010 TLVs and BEIs with 7th Edition Documentation. Cincinnati, OH: ACGIH; 2010.

51. Gorzinski SJ, Nolan RJ, Mc Collister SB, Kociba RJ, Mattsson JL. Subchronic oral toxicity, tissue distribution and clearance of hexachloroethane in the rat. *Drug Chem Toxicol.* 1985;8:155–69, <http://dx.doi.org/10.3109/01480548508999167>.
52. Gohlke R. [4,4-Diaminodiphenylmethane in a chronic experiment]. *Z Gesamte Hyg Ihre Grenzgeb.* 1978;24:159–62. German.
53. Dow Chemical U.S.A. Picloram. A two-year dietary chronic toxicity-oncogenicity study in Fisher 344 rats. Midland: Dow Chemical Company; 1986.
54. Bruckner JV, MacKenzie WF, Ramanathan R, Muralidhara S, Kim HJ. Oral toxicity of 1,2-dichloropropane: Acute, short-term, and long-term studies in rats. *Fundam Appl Toxicol.* 1989;12:713–30, [http://dx.doi.org/10.1016/0272-0590\(89\)90003-1](http://dx.doi.org/10.1016/0272-0590(89)90003-1).
55. Anderson RC. 90-Day subchronic oral toxicity in rats. Test material: Pyridine, Vol. 1. Pub. No. EPA-905/1-83-001; NTIS Pub. No. PB88-176136. Springfield: U.S. National Technical Information Service; 1987.
56. Buben JA, O'Flaherty EJ. Delinination of the role of metabolism in the hepatotoxicity of trichloroethylene and perchloroethylene: A dose-effect study. *Toxicol Appl Pharmacol.* 1985;78:105–22, [http://dx.doi.org/10.1016/0041-008X\(85\)90310-2](http://dx.doi.org/10.1016/0041-008X(85)90310-2).
57. Coate WB, Schoenfish WH, Lewis TR, Busey WM. Chronic inhalation exposure of rats, rabbits and monkeys to 1,2,4-trichlorobenzene. *Arch Environ Health.* 1977;32:249–55, <http://dx.doi.org/10.1080/00039896.1977.10667291>.
58. Reid JB. Saturated methyl halogenated aliphatic hydrocarbons. In: Bingham E, Cofrancesco J, Powell CH, editors. *Patty's toxicology*. 5th ed. Vol. 6. New York: John Wiley & Sons; 2001. p. 1–99, <http://dx.doi.org/10.1002/0471435139.tox062>.
59. Adams EM, Spencer HC, Rowe VK, McCollister BS, Irish DD. Vapor toxicity of trichloroethylene determined by experiments on laboratory animals. *Arch Ind Hyg Occup Med.* 1951;4:469–81.
60. Johannsen FR, Levinskas GJ, Rusch GM. Evaluation of the subchronic and reproductive effects of a series of chlorinated propanes in the rat. I. The toxicity of 1,2,3-trichloropropane. *J Toxicol Environ Health.* 1988;25:299–315, <http://dx.doi.org/10.1080/15287398809531211>.
61. Levine BS, Rust JH, Barkley JJ, Furedi EM, Lish PM. Six-month oral toxicity study on trinitrotoluene in beagle dogs. *Toxicology.* 1990;63:233–44, [http://dx.doi.org/10.1016/0300-483X\(90\)90045-I](http://dx.doi.org/10.1016/0300-483X(90)90045-I).
62. Lemen RA. Unsaturated halogenated hydrocarbons. In: Bingham E, Cofrancesco J, Powell CH, editors. *Patty's Toxicology*. 5th ed. Vol. 5. New York: John Wiley & Sons; 2001. p. 205–98.
63. Magnusson G, Bodin NO, Hansson E. Hepatic changes in dogs and rats induced by xylydine isomers. *Acta Pathol Microbiol Scand.* 1971;79:639–48.
64. Alexeeff GV, Broadwin R, Liaw J, Dawson SV. Characterization of the LOAEL-to-NOAEL uncertainty factor for mild adverse effects from acute inhalation exposures. *Regul Toxicol Pharmacol.* 2002;36:96–105, <http://dx.doi.org/10.1006/rtp.2002.1562>.
65. REACH. Reference preliminary Technical Guidance Document (reference p-TGD). Chapter 3, Human health hazard assessment. Helsinki: ECHA; 2011 [cited 2013 Sept 12]. Available from: <http://www.hse.gov.uk/aboutus/meetings/iacs/acts/watch/091106/p8annex1.pdf>.
66. DFG. Deutsche Forschungsgemeinschaft. List of MAK and BAT Values 2010. Report No. 46. Weinheim: Wiley-VCH Verlag GmbH & Co. KGaA; 2010.
67. CIOP-PIB. Interdepartmental commission for maximum admissible concentrations and intensities for agents harmful to health in the working environment. Harmful substances in occupational environment, admissible concentrations. Warszawa: CIOP-PIB; 2010.
68. Simon-Hettich B, Rothfuss A, Steger-Hartmann T. Use of computer-assisted prediction of toxic effects of chemical substances. *Toxicology.* 2006;224:156–62, <http://dx.doi.org/10.1016/j.tox.2006.04.032>.